Sequencing Technologies – Overview of PCR and Sanger method of DNA sequencing



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ABOUT INQABABIOTEC















Our Mission: Catalysing Africa's Prosperity Through Genomics!

Core Services











DNA SYNTHESIS (PRIMERS & PROBES)

SANGER DNA SEQUENCING

SNP GENOTYPING & METHYLATION ANALYSIS

qPCR ANALYSIS







BIOINFORMATICS SOLUTIONS



ANIMAL GENETICS



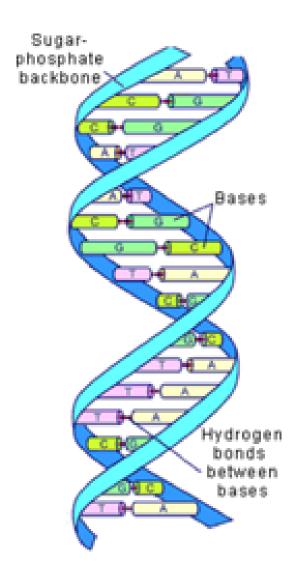
INSTRUMENTS SERVICE, REPAIRS & MAINTENANCE



TECHNICAL SESSIONS

Polymerase Chain Reaction



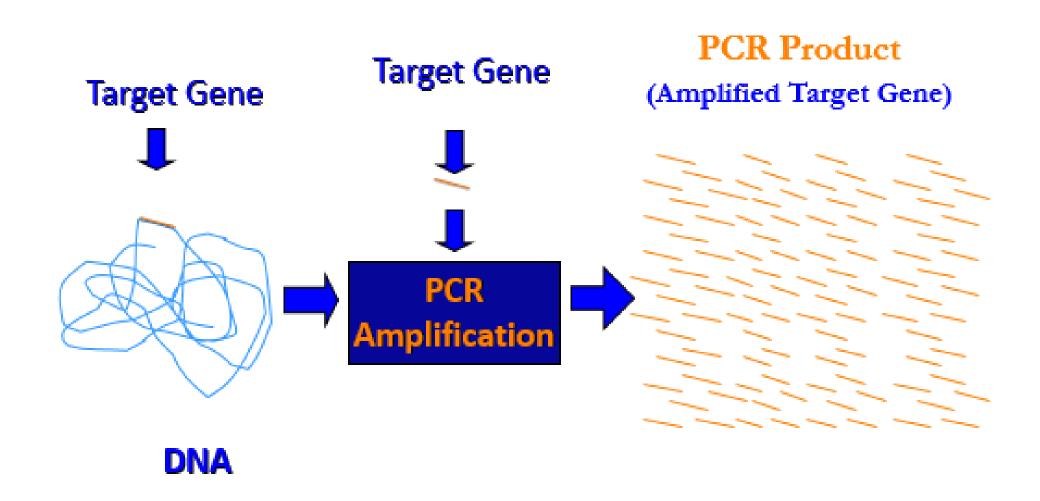


Polymerase Chain Reaction (PCR) refers to an in-vitro technique which is nucleic- acid based for making numerous copies of a DNA fragment of interest, such as a gene for its identification and characterization. For an example: HPV, BBTV, YMV, insect identification etc.

An in-vivo technique use to make more copies of a virus or any other gene require the use of living organisms such as bacteria in a procedure called cloning and transformation.

Polymerase Chain Reaction





Mechanism

Polymerase Chain Reaction



During PCR, a thermocycler brings the reaction mix to 3 different temperatures analogous to the 3 steps of DNA replication

Denaturation (94°C) of the DNA template by heat Annealing (37°-70°C) of the primers to the template Extension (72°C) of the DNA strand by DNA polymerase



These steps are repeated for 25 to 40 cycles

Denaturation



Denaturation occurs at 94°C

The high temperature is used to break down the hydrogen bonds that hold the two strands together



Annealing



Annealing occurs at 37°-70°C

Oligonucleotide DNA primers anneal to their complementary sequences on the template strands Annealing temperature depends on the melting temperature (T_m) of the primer (dependent on base composition)

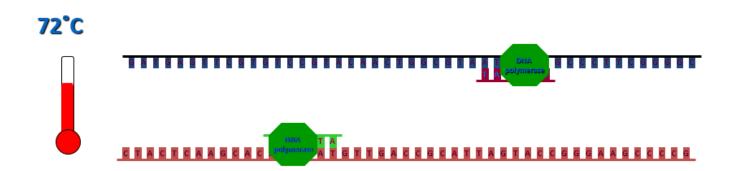


Extension



Extension occurs at 72°C DNA polymerase attaches to the primers and extends the new DNA strand

The 3 steps (denaturation, annealing, and extension) are repeated for another 24 to 29 cycles





Target Sequence

A desired target sequence is identified

To isolate the target sequence, primers that flank the region must be constructed

The DNA segment that is then amplified contains the region of interest

Template DNA

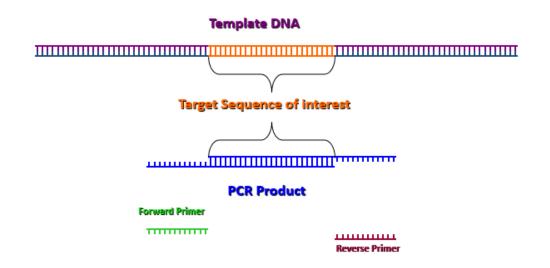
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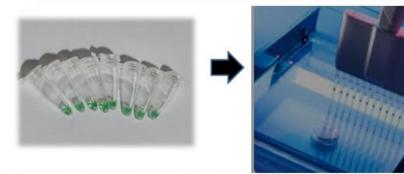


Post-PCR Assay

Gel electrophoresis



Purify PCR products, run on agarose gel electrophoresis, take gel picture under UV light and subject to sanger DNA sequencing



1500 bp- 16s DNA fragment from bacteria cultures resolved on 1.5% gel

Method of gel electrophoresis

Melt agarose powder in buffer, cool and add DNA stain (Safe Red). Mix thoroughly

Pour into casting tray with comb and allow to solidify

Add running buffer to the gel tank, place gel tray in and load DNA samples and marker View DNA on UV light box and show results

Run gel at constant voltage until band separation occurs



480-720 bp- ITs DNA fragment from Fungi isolates resolved on 2% gel

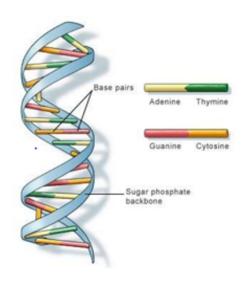


SEQUENCING TECHNOLOGIES

Sequencing Technologies



Determines the order of nucleotide (G, A, T, and C) in a stretch of DNA



- Sanger Sequencing
- Illumina Sequencing: MiSeq, HiSeq and PacBios
- Nanopore, MinION, Single-molecule

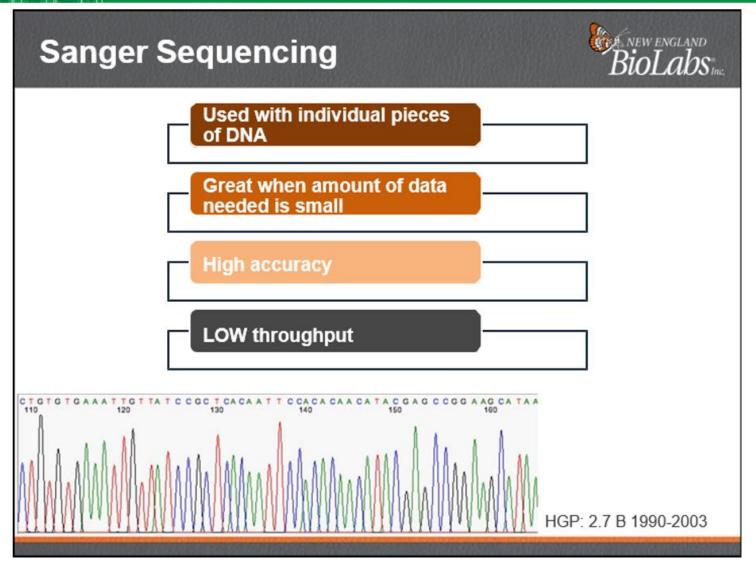
Application of Sequencing







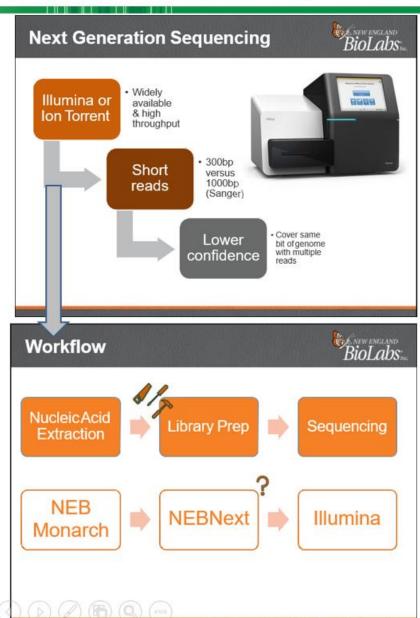


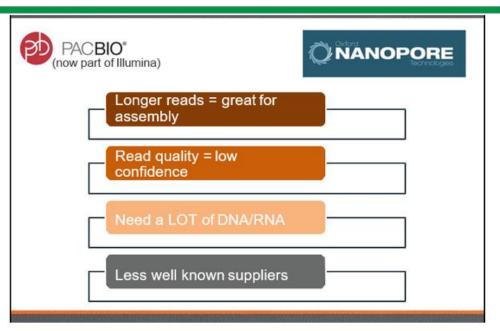


Human genome was completed using Sanger at the cost of 2.7 billion US dollars. It took from 1990 to 2003. NGS the cost is now 1 K \$

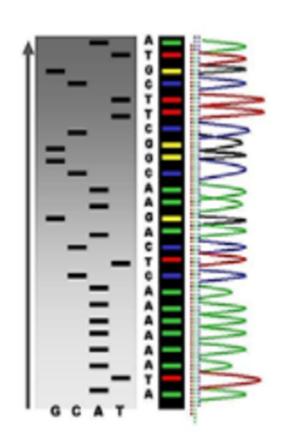












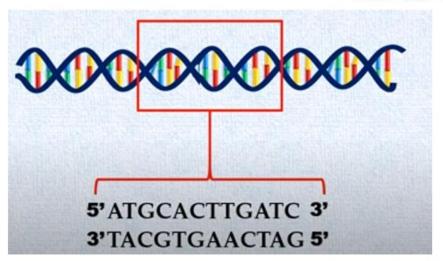


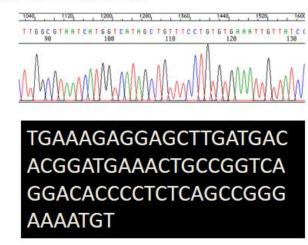
Sequencing Technologies



DNA Sequencing-

It is a technique whereby precise order of nucleotides (G, A, T, and C) in a stretch of DNA can be determined





Developed by Fred Sanger in 1977 using chemically altered "dideoxy" bases to terminate newly synthesized DNA fragments at specific bases (either A, C, T, or G). These fragments are then size-separated, and the DNA sequence can be read.

An unknown sequence can be matched up to known sequences published in the Database:

List all sequences producing significant alignments

Gene identification

Organism genus/species

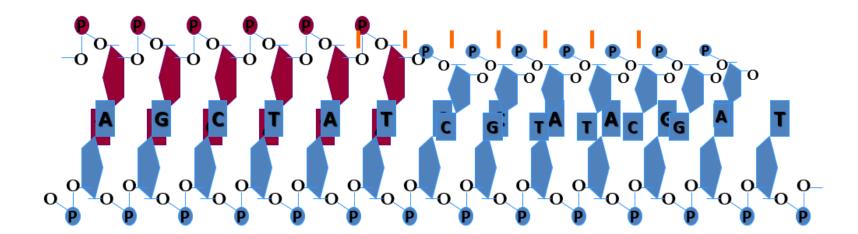
Identity alignment/match





Modern Sanger sequencing is based on the principles of Di-deoxynucleotides in DNA Polymerization

- 1) DNA Template
- 2) Primer
- 3) DNA Polymerase
- 4) dNTPs
- 5) Mg²⁺ ions



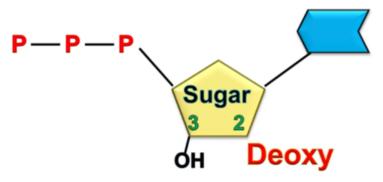
Principles of Sanger Sequencing



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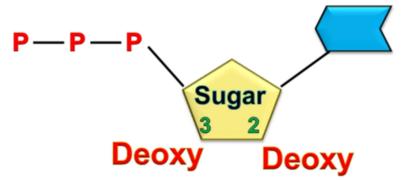


If the 3'OH group of the nucleotide is absent, then there wont be addition of new nucleotides and the reaction would stop. The incorporation of dideoxy nucleotide and termination of polymerization reaction is a random process. Hence the overall mixture will have several fragements that have experienced the chain termination. If the length of each fragment is known then the location of nucleotides in the DNA can be estimated



Nucleotide

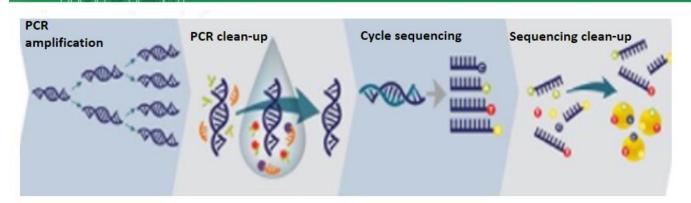
Deoxynucleotide- present in DNA

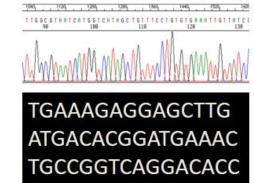


Di- deoxynucleotide

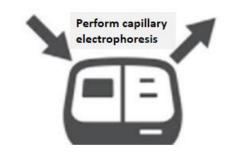
Sanger Sequencing Workflow

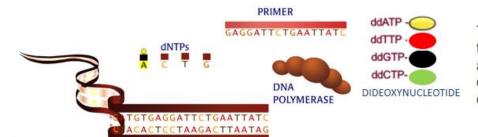






In order to determine the sequence, Sanger sequencing makes use of chemical analogs of the four nucleotides in DNA. These analogs, called dideoxyribonucleotides (ddNTPs), are missing the 3' hydroxyl group that is required for 5' to 3' extension of a DNA polynucleotide chain. By mixing ddNTPs that have been labeled with a different color for each base, unlabeled dNTPs, and template DNA in a polymerase-driven reaction, strands of each possible length are produced when the ddNTPs are randomly incorporated and terminate the chain.



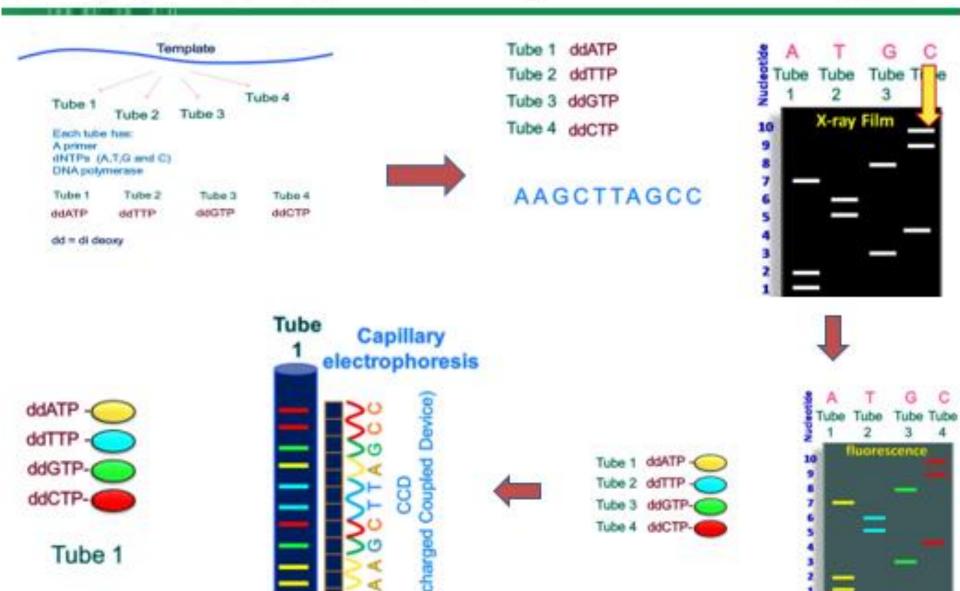


TEMPLATE DNA

The extension products are then separated by electrophoresis, resolved to single-nucleotide differences in size. The chain-terminated fragments are detected by their fluorescent labels, with each color identifying one of the terminating ddNTPs. The sequence of the template DNA strand can thus be derived by analysis

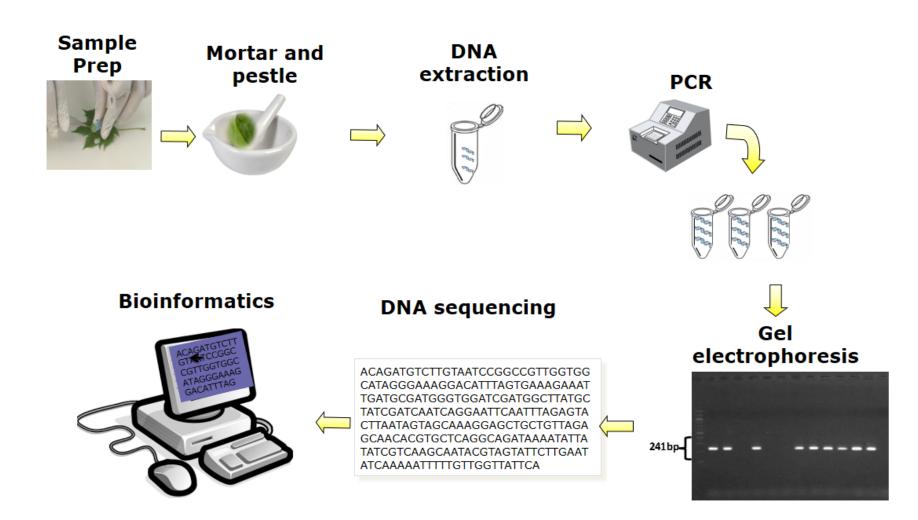
Advancement in Sanger Sequencing





Technical Review





Technical Precautions in Sequencing



√ Poor Template Quality

Poor template quality is the most common cause of sequencing problems characterized by:

- Noisy data or peaks under peaks
- · No usable sequence data
- Weak signal

Always follow recommended procedures to prepare templates.

✓ Contamination

Potential contaminants include:

- Proteins
- RNA
- Chromosomal DNA

✓ Excessive Reagents

Excess PCR primers, dNTPs, enzyme, and buffer components (from a PCR amplification used to generate the sequencing template)

- Residual salts
- Residual organic chemicals such as phenol, chloroform, and ethanol
- Residual detergents



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Sample Storage Devices



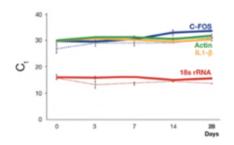
Collection Devices

DNA/RNA Shield™ Sample Collection and Preservation



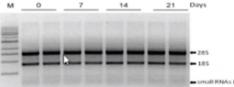


RNA Stabilization and Storage



RNA in blood is effectively stabilized in DNA/RNA Shield $^{\text{IM}}$ at ambient temperature.

RNA was purified from whole blood stored for the indicated time points in DNA/RNA Shield[™] and analyzed by RT-qPCR. Solid and dotted lines represent two different donors.



Cellular RNA is effectively stabilized in DNA/RNA Shield $^{\text{TM}}$ at ambient temperature.

RNA was purified from cells (HCT 116) stored in DNA/RNA Shield™ at the designated times.

■ small RNAs (inc. miRNA)







Sanger Sequencing

Advantages	Disadvantages
Lowest error rate	High cost per base
Long read length (~750 bp)	Long time to generate data
Can target a primer	Need for cloning
	Amount of data per run

Illumina Sequencing

Advantages	Disadvantages
Low error rate	Must run at very large scale
Lowest cost per base	Short read length (50-75 bp)
Tons of data	Runs take multiple days
	High startup costs
	De Novo assembly difficult

PacBio Sequencing

Advantages	Disadvantages
Can use single molecule as template	High error rate (~10-15%)
Potential for very long reads (several kb+)	Medium/high cost per base
	High startup costs



Final Thoughts

- DNA sequencing is becoming vastly faster and more affordable
- Generating data is no longer the bottleneck, understanding it is
- Bioinformatics types should be in high demand in the near future





•An ergonomic laboratory chair is a great option for people who need to slide around a space, whether it is because they have to work on multiple samples at once or because they need to provide instructions to the rest of the members of the staff

Microscopes

Ensure that you can view the eyepiece while sitting or standing in an upright position. This includes the shoulders, back and neck

 Hoods and Biological Safety Cabinets Keep arms relaxed and by the sides. Back, shoulders and neck should be upright and neutral in position avoiding an awkard position.









ALWAYS ON HAND AS A LAB STOOL.

The Labsit lab stool is on hand wherever you need it. It is the uncomplicated solution for short periods of sitting. However the Labsit lab stool is in no way inferior to the swivel chair when it comes to laboratory suitability. It is washable, resistant to disinfectants and robust. The PU-foam upholstery is very comfortable.







BRINGS COLOUR TO THE LABORATORY.















THANK YOU

